

Vibrational Spectroscopic & Molecular Docking Studies of 2,6-Dichlorobenzyl Alcohol

Merrin Mary Abraham^a, Resmi K. S.^a, Sheena Mary Y.^b, C. YohannanPanicker^{b*}, B. Harikumar^c ^aDepartment of Physics, TKM College of Arts and Science, Kollam, Kerala, India ^bDepartment of Physics, Fatima Mata National College, Kollam, Kerala, India ^cDepartment of Chemistry, TKM College of Arts and Science, Kollam, Kerala, India (**E-mail: sypanicker@rediffmail.com*)

Abstract: The optimized molecular structure, vibrational frequencies, and corresponding vibrational assignments of 2,6dichlorobenzyl alcohol have been investigated experimentally and theoretically using Gaussian09 software package. Potential energy distribution of the normal modes of vibrations was done using GAR2PED program. The HOMO and LUMO analysis was used to determine the charge transfer with the molecule. The stability of the molecule arising from hyper-conjugative interaction and charge delocalization has been analyzed using NBO analysis. Molecular electrostatic potential was performed by the DFT method and from the MEP, it is evident that the negative region covers the CH₂ group, oxygen atom, and phenyl ring and that positive region is over the hydrogen atoms. The calculated first hyperpolarizability of the title compound is 4.523 times that of standard NLO material urea and the title compound is an attractive object for future studies of nonlinear optical properties. The docked title compound forms a stable complex with aryl hydrocarbon receptor and gives a binding affinity value of -4.4 kcal/mol. The results suggest that the compound might exhibit inhibitory activity against aryl hydrocarbon receptor.

Key Words: DFT, chlorobenzyl, docking, NLO, MEP.

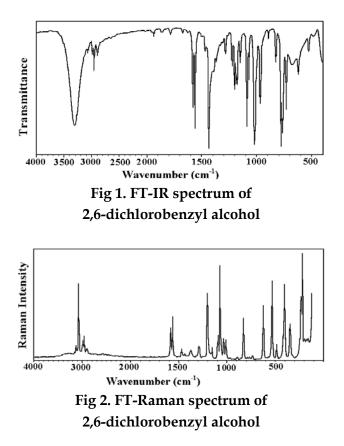
INTRODUCTION

Benzyl alcohol derivatives are found in natural products and play a central role in numerous mechanistic investigations [1]. Aminobenzyl alcohols are useful as antimicrobial agents [2] and herbicides [3]. 3-Aminobenzyl alcohol is used to synthesize gamma-L-glutamyl-4nitroanilide derivative to determine y-GTP (gammaglutamyltranspeptides) in serum [4]. A hit-to-lead optimization program on dichlorobenzyl derivative discovers pyrimidine-5-carbonitrile-6-cyclopropyl as a functional antagonist of the human CXCR2 receptor and shows good oral bioavailability in the rat [5]. Alcohols are used in topical ophthalmic pharmaceuticals and are useful against cataracts [6]. In spite of these numerous applications and consequent interest in their qualitative and quantitative characterization, the vibrational spectra of benzyl alcohol derivative provide a deeper insight into their biological actions when they are administered as drugs and in the environment as herbicides. Several author groups have studied the vibrational spectra of benzyl alcohol derivatives [7, 8].

EXPERIMENTAL & COMPUTATIONAL DETAILS

The FT-IR spectrum (Fig. 1) was recorded using KBr pellets on a DR/Jasco FT-IR 6300 spectrometer and the FT-Raman spectrum (Fig. 2) was obtained on Bruker RFS 100/s, Germany.

Calculations of the title compound were carried out with Gaussian09 [9] program using the B3LYP/ 6-31G (6D, 7F) basis sets to predict the molecular structure and vibrational wave numbers. Molecular geometry was fully optimized by Berny's optimization algorithm using redundant internal coordinates. Harmonic vibrational wave numbers were calculated using the analytic second derivatives to confirm the convergence to minima on the potential surface. The theoretical calculations were performed with the hybrid B3LYP functional that is, a combination of the Becke's three-parameter exchange functional and Lee-Yang-Parr correlation functional [10, 11]. The DFT calculations reported excellent vibrational wavenumber of organic compounds if the calculated wavenumbers were scaled to compensate for the approximate treatment of electron correlation, for basis set deficiencies and for the anharmonicity [12]. The DFT hybrid B3LYP functional tended to overestimate the fundamental modes [13]; therefore, scaling factor of 0.9613 has to be used for obtaining a considerably better agreement with experimental data [14]. Then, frequency calculations were employed to confirm the structure as minimum points in energy. The absence of imaginary wavenumbers on the calculated vibrational spectrum confirmed that the structure (Fig. 3) deduced corresponded to minimum energy. The assignments of the calculated wave numbers were aided by the animation option of GAUSSVIEW program, which gave a visual presentation of the vibrational modes [15]. The potential energy distribution (PED) was calculated with the help of GAR2PED software package [16].



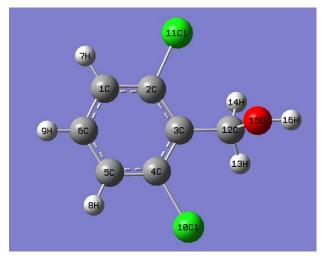


Fig 3. Optimized geometry of 2,6-dichlorobenzyl alcohol

RESULTS & DISCUSSION

IR and Raman Spectrum

The calculated (scaled) wavenumbers, observed IR, Raman bands, and assignments are given in Table 1. Aromatic compounds commonly exhibit multiple weak bands in the region 3100-3000 cm⁻¹, due to aromatic CH stretching vibrations [17]. For the title compound, the band observed at 3094 cm-1 in the Raman spectrum and at 3127, 3121, 3095 cm⁻¹ (DFT) were assigned the CH stretching modes of the phenyl ring. For tri-substituted benzenes δCH modes were expected in the range 1050-1280 cm⁻¹ [17] and the bands observed at 1214 cm-1in the IR spectrum, 1071 cm⁻¹in the Raman spectrum and at 1230, 1184, 1066 cm⁻¹ (DFT) are assigned as these in-plane CH deformation modes. The CH out-of-plane deformations were expected below 1000 cm⁻¹ [18]. The bands at 971, 900 cm-1in the IR spectrum, 900 cm-1in the Raman spectrum and 980, 947, 909 cm⁻¹ (DFT) were assigned as the CH outof-plane deformations.

The benzene ring possesses six ring stretching modes of which the four with the highest wavenumbers occurring near 1600, 1580, 1490, and 1440 cm⁻¹ were good group vibrations [17]. These modes were expected in the region 1250-1620 cm⁻¹ [17]. For the title compound, the bands observed at 1571, 1557, 1429 cm⁻¹ (IR), 1578, 1437, 1294 cm⁻¹ (Raman) and 1585, 1552, 1438, 1437, 1310 cm⁻¹ (DFT) were assigned as the phenyl ring stretching modes.

B3LYP/6-31G*		IR	Raman			
v(cm ⁻¹)	IR _A	R _A	υ(cm ⁻¹)	υ(cm-1)	Assignments ^a	
3490	4.97	263.88	3300		vOH(100)	
3127	3.95	175.44			vCH(98)	
3121	0.70	64.76			υCH(95)	
3095	5.56	84.57		3094	υCH(97)	
2987	17.88	40.62		3000	υCH2(100)	
2946	34.69	79.83	2943		vCH2(93)	
1585	29.67	23.88	1571	1578	υPh(65), δCH2(12)	
1552	45.74	13.81	1557		υPh(70), δCH2(10)	
1507	5.64	12.68			δCH2(49), υPh(12)	
1438	1.30	0.83			υPh(54), δCH2(20)	
1437	64.38	0.08	1429	1437	υPh(60), δOH(22)	
1413	15.12	8.69			δOH(45), vPh(18)	
1310	0.61	11.89		1294	υPh(59), δCH2(23)	
1233	1.23	1.21			δCH2(55)	
1230	20.45	13.24	1214		δCH(71), δCH2(11	
1184	11.01	7.23			δCH(63), υCC(21)	
1163	50.42	13.48		1167	υCO(41), δCH(13)	
1156	14.89	0.90	1157		δCH2(58), υCC(15)	
1066	4.02	18.81		1071	δCH(45), δCH2(18	
1049	30.61	7.17	1057	1040	υCC(39), δCH(23	
984	22.45	3.12			δCH2(61), υCC(12)	
980	0.99	0.30	971		γCH(95)	
947	60.08	14.78			υCO(44), γCH(34)	
909	0.00	0.77	900	900	γCH(68)	
808	8.06	2.83	829	794	υCCl(35), γCH(23	
788	32.93	3.15	786	778	υCCl(41), γCH(17)	
745	39.08	1.86		754	τPh(55), υCCl(24)	
720	90.03	0.13	729		υPh(48), υCCl(20)	
599	4.54	5.71			γОН(37), τСН2(18)	
533	0.70	8.52		540	τPh(40), γOH(22)	
531	0.04	1.45	514		τPh(37), γOH(16)	
473	4.24	0.25		484	484 τCH2(48), τPh(30	
384	9.18	6.53		δPh(32), γCCl(2		
381	10.21	7.69		τPh(40), γCCl(2		
325	0.99	5.75		δPh(38), γCCl		
307	2.59	0.06			τCH2(29), γCCl(22	
211	3.15	4.07		219	δCCl(24), δPh(19)	
210	137.83	8.10		200	δCCl(28), δPh(25)	
196	1.46	1.79			τPh(38)	
162	3.44	2.11		141	τPh(31)	
77	2.87	1.37			τCCl(24), τPh(19)	
37	1.17	2.63			τOH(27), τPh(20)	

Table 1: Vibrational assignments of 2,6-dichlorobenzyl alcohol

υ- stretching; δ- in-plane deformation; γ- out-of-plane deformation; ^a% of PED contribution of each mode is given in parenthesis.

In asymmetric tri-substituted benzenes, when all the three substituents were light, the wavenumber interval of the breathing mode was between 500 and 600 cm⁻¹. In the case of mixed substituent, the wavenumber was expected to appear between 600 and 750 cm⁻¹. When all the three substituents were heavy, the ring breathing mode was expected around 1100 cm⁻¹ [19, 20]. The band observed at 729 cm⁻¹in the IR spectrum was assigned as the ring breathing mode of the phenyl ring which found support from the computational value at 720 cm⁻¹.

The asymmetric and symmetric CH₂ stretching appeared in the region 3000±50 and 2965±30 cm⁻¹, respectively [17, 18]. The CH₂ stretching modes were observed at 2943 cm⁻¹ in the IR spectrum and at 3000 cm⁻¹ in the Raman spectrum. The DFT calculations gave these modes at 2987 and 2946 cm⁻¹. The scissoring vibration δ CH₂ and wagging vibration ω CH₂ appeared in the regions 1455 ± 55 and 1350 ± 85 cm⁻¹, respectively [17,18]. The CH₂ deformation band, which came near 1463 cm⁻¹ in alkenes, [21] was lowered to about 1440 cm⁻¹ when the CH₂ group was next to a double or triple bond. The rocking mode [17] ρ CH2 was expected in the range 895±85 cm⁻¹. The deformation modes of the methylene group were assigned at 1507 cm⁻¹ (scissoring), 1233 cm⁻¹ (wagging), 1156 cm⁻¹ (twisting), and 984 cm⁻¹ (rocking) theoretically.

For the hydroxyl group, the OH group provided three normal vibrations; the stretching vibration OH, in-plane and out-of-plane deformations δ OH and γ OH. The inplane OH deformation [17] was expected in the region 1440 ± 40 cm⁻¹. The out-of-plane deformation was expected generally in the region 650 ± 80 cm⁻¹ [17]. The C-O stretching mode was expected in the region 1220 ± 40 cm⁻¹ [18-20]. The OH modes were assigned at 3490 cm⁻¹ (stretching), 1413 cm⁻¹ (in-plane bend), and 599 cm⁻¹ (outof-plane bend) theoretically, for the title compound.

Varghese et al. reported vOH at 3633 cm⁻¹ and δ OH at 1345 cm⁻¹ theoretically and C-O stretching at 1255 cm⁻¹ in both IR and Raman spectra and 1262 cm⁻¹ theoretically [22]. For paracetamol, the C-O stretching mode and out-of-plane OH were reported at 1240 and 620 cm⁻¹, respectively [23]. In the present case the C-O stretching mode was assigned at 1167 cm⁻¹ in the Raman spectrum and at 1163 cm⁻¹ theoretically. For simple organic chlorine compounds C-Cl absorptions were in the region 800-700 cm⁻¹ [24, 25]. The bands observed at 829, 786 cm⁻¹ in the IR spectrum, 794, 778 cm⁻¹ in the Raman spectrum and at 808, 788 cm⁻¹ (DFT) were assigned as the C-Cl stretching modes for the title compound. Most of the modes were not pure but contained significant contributions from other modes also.

Nonlinear optical properties

The first hyperpolarizability (β_0) of this novel molecular system is calculated using the B3LYP/ 6-31G (6D, 7F) method, based on the finite-field approach. In the presence of an applied electric field, the energy of a system is a function of the electric field. The first hyperpolarizability is a third-rank tensor that can be described by a 3 × 3 × 3 matrix. The 27 components of the 3D matrix can be reduced to 10 components due to the Kleinman symmetry [26]. The components of β are defined as the coefficients in the Taylor series expansion of the energy in the external electric field. When the electric field is weak and homogeneous, this expansion becomes:

$$E = E_0 - \sum_i \mu_i F^i - \frac{1}{2} \sum_{ij} \alpha_{ij} F^i F^j - \frac{1}{6} \sum_{ijk} \beta_{ijk} F^i F^j F^k - \frac{1}{24} \sum_{ijkl} \gamma_{ijkl} F^i F^j F^k F^l + \dots$$

where E_0 is the energy of the unperturbed molecule, F^i is the field at the origin, μ_i , α_{ij} , β_{ijk} and γ_{ijkl} are the components of dipole moment, polarizability, the first hyperpolarizabilities, and second hyperpolarizabilities, respectively.

$$\beta_0 = (\beta_x^2 + \beta_y^2 + \beta_z^2)^{1/2}$$

where
$$\beta_x = \beta_{xxx} + \beta_{xyy} + \beta_{xzz}$$

$$\beta_y = \beta_{yyy} + \beta_{xxy} + \beta_{yzz}$$

$$\beta_z = \beta_{zzz} + \beta_{xxz} + \beta_{yyz}$$

The calculated first hyperpolarizability of the title compound is 0.588×10^{-30} esu which is 4.523 times that of standard NLO material urea (0.13 ×10⁻³⁰ esu) [27]. We conclude that the title compound is an attractive object for future studies of nonlinear optical properties.

Molecular Electrostatic Potential (MEP)

MEP is related to the Electron Density (ED) and is a very useful descriptor in understanding sites for electrophilic and nucleophilic reactions, as well as hydrogen bonding interactions [28]. The electrostatic potential V(r) is also well suited for analyzing processes based on the "recognition" of one molecule by another, as in drug-receptor and enzyme-substrate interactions, because it is through their potentials that the two species first "see" each other [29,30]. To predict reactive sites of electrophilic and nucleophilic attacks for the investigated molecule, MEP at the B3LYP/ 6-31G (6D, 7F) optimized

geometry is calculated. The different values of the electrostatic potential at the surface are represented by different colors: red represents regions of most electronegative, electrostatic potential; blue represents regions of the most positive electrostatic potential; and green represents region of zero potential. Potential decreases in the order red < orange < yellow < green < blue. The MEP surface (Fig.4) provides necessary information about the reactive sites. From the MEP, it is evident that the negative region covers the CH₂ group, oxygen atom, phenyl ring, and the positive region is over the hydrogen atoms.

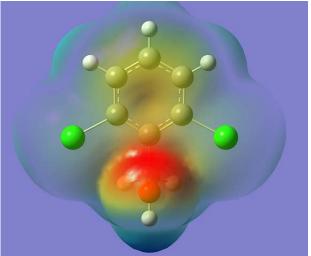
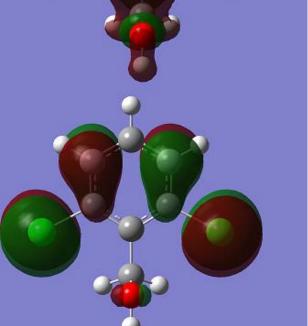


Fig 4. MEP plot of 2,6-dichlorobenzyl alcohol

Frontier Molecular Orbital Analysis

The most widely used theory by chemists is the molecular orbital (MO) theory. It is important that potential electron affinity ionization (I), (A), electrophilicity index (ω), chemical potential (μ), electronegativity (χ) , and hardness (η) be put into a MO framework. Based on density functional descriptors, global chemical reactivity descriptors of compounds, such as hardness, chemical potential, softness, electro negativity, and electrophilicity index, as well as local reactivity, have been defined [31-33]. Pauling introduced the concept of electro-negativity as the power of an atom in a compound to attract electrons to it. Using Koopman's theorem for closed shell components η , μ and χ can be defined as $\eta = (I - A)/2$; $\mu = -(I + A)/2$; $\chi = (I + A)/2$; where I and A are the ionization potential and electron affinity of the compounds, respectively. The ionization energy (I) and electron affinity (A) can be expressed through HOMO and LUMO orbital energies as I = $-E_{HOMO}$ = 8.086 and A =



 $-E_{LUMO} = 4.413$ eV. Electron affinity refers to the capability

of ligand to accept precisely one electron from a donor.

However, in many kinds of bonding viz. covalent

hydrogen bonding, partial charge transfer takes place.

Considering the chemical hardness (η) , a large HOMO-

LUMO energy gap means a hard molecule and a small

HOMO-LUMO gap means a soft molecule.

Fig 5. HOMO-LUMO plots of 2,6-dichlorobenzyl alcohol

One can also relate the stability of the molecule to hardness, which means that the molecule with a smaller HOMO-LUMO gap (3.673eV) is more reactive. Parr et al. [31] have defined a new descriptor to quantify the global electrophilic power of the compound as electrophilicity index (ω) which defines a quantitative classification of global electrophilic nature of a compound. Parr et al. have proposed electrophilicity index (ω) as a measure of energy lowering due to maximal electron flow between donor and acceptor. They defined electrophilicity index as follows: ω = $\mu^2/2\eta$. The usefulness of this new reactivity measure has been recently demonstrated in understanding the toxicity of various pollutants in terms of their reactivity and site selectivity [34]. The calculated values of ω , μ , χ , and η are 10.6324eV, -6.250eV, 6.250eV, and 1.837eV, respectively. The calculated value of electrophilicity index describes the biological activity of the title compound. The atomic orbital components of the frontier molecular orbital are shown in Fig. 5.

Natural Bond Orbital Analysis

The natural bond orbital (NBO) calculations were performed using NBO 3.1 program [35] as implemented in the Gaussian09 package at the DFT/B3LYP/ 6-31G (6D, 7F) level in order to understand various second-order interactions between the filled orbital of one subsystem and the vacant orbital of another subsystem, which is a measure of the intermolecular delocalization or hyperconjugation. NBO analysis provides the most accurate possible 'natural Lewis structure' picture of 'j' because all orbital details are mathematically chosen to include the highest possible percentage of the electron density. A useful aspect of the NBO method is that it gives information about interactions of both filled and virtual orbital spaces that could enhance the analysis of intra- and inter-molecular interactions. The second-order Fockmatrix was carried out to evaluate the donor-acceptor interactions in the NBO basis. The interactions resulted in a loss of occupancy from the localized NBO of the idealized Lewis structure into an empty non-Lewis orbital. For each donor (i) and acceptor (j) the stabilization energy (E2) associated with the delocalization $i \rightarrow j$ is determined as:

E (2) =
$$\Delta E_{ij} = q_i \frac{(F_{i,j})^2}{(E_i - E_i)}$$

 q_i is donor orbital occupancy, E_i , E_j is the diagonal elements, and F(i,j) is the off diagonal NBO Fock- matrix element. In NBO analysis large E(2) value shows the intensive interaction between electron-donors and electron- acceptors, and a higher extension of conjugation of the whole system. The possible intensive interaction is given in Table 2. The second-order perturbation theory analysis of Fock-matrix in NBO basis shows that strong

intra-molecular, hyper-conjugative interactions are formed by orbital overlap between n(Cl) and $\pi^*(C-C)$ bond orbital which result in ICT causing stabilization of the system. These interactions are observed as an increase in electron density(ED) in C-C anti-bonding orbital that weakens the respective bonds. The strong intra-molecular, hyper-conjugative interaction of C₃-C₄ from Cl₁₀ of n₃(Cl₁₀) $\rightarrow \pi^*(C_3-C_4)$ which increases ED(0.39326e) that weakens the respective bonds C₃-C₄ leading to stabilization of 9.63kJ/mol and also the hyper-conjugative interaction of C₁-C₂ from Cl₁₁ of n₃(Cl₁₁) $\rightarrow \pi^*(C_1-C_2)$ which increases ED (0.37193e) that weakens the respective bonds C₁-C₂ leading to stabilization of 9.29kJ/mol.

The NBO analysis describes the bonding in terms of the natural hybrid orbital n₃(Cl₁₀), which occupies a higher energy orbital (-0.31889a.u) with considerable p-character (100%) and low occupation number (1.94196) and the other n₁(Cl₁₀) occupies a lower energy orbital (-0.92091a.u) with p-character (15.25%) and high occupation number (1.99411). The NBO analysis also describes the bonding in terms of the natural hybrid orbital n₃(Cl₁₁), which occupies a higher energy orbital (-0.31888a.u) with considerable pcharacter (100%) and low occupation number (1.94195) and the other n1(Cl11) occupies a lower energy orbital (-0.92090a.u) with p-character (15.25%) and high occupation number (1.99412). Again, the NBO analysis describes the bonding in terms of the natural hybrid orbital $n_2(O_{15})$, which occupies a higher energy orbital (-0.28535a.u) with considerable p-character (100%) and low occupation number (1.96816) and the other $n_1(O_{15})$ occupies a lower energy orbital (-0.60657a.u) with p-character (46.89%) and high occupation number (1.98601). Thus, a very close to pure p-type lone pair orbital participates in the electron donation to the $\pi^*(C_3-C_4)$ orbital for $n_3(Cl_{10}) \rightarrow \pi^*(C_3-C_4)$ and $\pi^*(C_1-C_2)$ orbital for $n_3(Cl_{11}) \rightarrow \pi^*(C_1-C_2)$ interaction in the compound. The results are tabulated in Table 3.

Molecular docking

Aryl hydrocarbon receptor (AHR), a cytosolic ligandactivated transcription factor, belongs to the family of hetero-dimeric transcriptional regulators and is widely expressed in a variety of animal and human species, and experimental animal data provided substantial support for an association between abnormal AHR function and cancer, implicating AHR may be a novel drug-interfering target for cancers [36]. Certain benzyl alcohol derivatives show anticancer activity [37]. High-resolution crystal

Donor(i)	Type	ED/e	Acceptor(j)	Type	ED/e	E(2)a	E(j)-E(i)b	F(i,j)c
C1-C2	σ	1.98163	C1-C6	σ*	0.01622	2.34	1.29	0.049
			C2-C3	σ*	0.03356	3.98	1.28	0.064
			C3-C12	σ*	0.02501	2.94	1.14	0.052
C1-C2	п	1.67720	C3-C4	π*	0.39326	20.04	0.29	0.069
			C5-C6	π*	0.32742	19.19	0.29	0.067
C3-C4	σ	1.97002	C2-C3	σ*	0.03356	3.49	1.27	0.060
			C2-Cl11	σ*	0.03486	4.69	0.82	0.056
			C3-C12	σ*	0.02501	2.37	1.14	0.046
			C4-C5	σ*	0.02172	3.51	1.29	0.060
C3-C4	п	1.66492	C1-C2	π*	0.37193	19.41	0.28	0.067
			C5-C6	π*	0.32742	19.58	0.29	0.068
			C12-O15	σ*	0.02140	5.26	0.53	0.051
C2-C3	σ	1.97002	C1-C2	σ*	0.02172	3.51	1.29	0.060
			C3-C4	σ*	0.03357	3.49	1.27	0.060
			C4-Cl10	σ*	0.03486	4.69	0.82	0.055
C4-C5	σ	1.98163	C3-C4	σ*	0.03357	3.98	1.28	0.064
			C3-C12	σ*	0.02501	2.94	1.14	0.052
			C5-C6	σ*	0.01622	2.34	1.29	0.049
LPCl10	σ	1.99411	C3-C4	σ*	0.03357	1.02	1.46	0.059
LPCl10	п	1.97381	C3-C4	σ*	0.03357	3.46	0.86	0.049
			C4-C5	σ*	0.02172	2.56	0.87	0.042
LPC110	n	1.94196	C3-C4	π*	0.39326	9.63	0.33	0.055
LPCl11	σ	1.99412	C2-C3	σ*	0.03356	1.02	1.46	0.035
LPCl11	п	1.97382	C1-C2	σ*	0.02172	2.56	0.87	0.042
			C2-C3	σ*	0.03356	3.46	0.86	0.049
LPCl11	n	1.94195	C1-C2	π*	0.37193	9.29	0.32	0.053

Table 2. TG-DTA results evidenced the presence of lattice water in Co(II) and Ni(II) complexes

^a E(2) means energy of hyper-conjugative interactions (stabilization energy in kJ/mol)

^b Energy difference (a.u) between donor and acceptor i and j NBO orbitals ^c F(i,j) is the Fock matrix elements (a.u) between i and j NBO orbitals

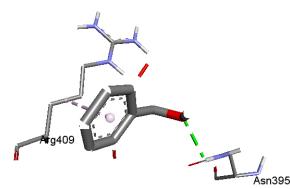
Bond(A-B)	ED/eª	EDA%	EDB%	NBO	s%	p%
σC1-C2	1.98163	49.23	50.77	0.7016(sp1.90)C	34.35	65.65
001-02	-0.73810	17.20	00	+0.7126(sp1.51)C	39.75	60.65
πC1-C2	1.67720	48.14	51.86	0.6938(sp1.00)C	0.00	100.0
101-02	-0.27880	10.11		+0.6938(sp1.00)C	0.00	100.0
σ C3-C4	1.97002	50.52	49.48	0.7108(sp1.95)C	33.86	66.14
0C3-C4	-0.73425	50.52		+0.7034(sp1.95)C	39.55	60.45
	1.66492	48.44	51.56	0.6960(sp1.00)C	0.01	99.99
πC3-C4	-0.27670	40.44		+0.7180(sp1.00)C	0.00	100.0
	1.98163	E0 77	49.23	0.7126(sp1.51)C	9.75	60.25
σC4-C5	-0.73811	50.77	49.23	+0.7016(sp1.91)C	34.35	65.65
10110	1.99411			sp0.18	84.75	15.25
n1Cl10	-0.92091					
2 C110	1.97381				0.05	00.05
n2 Cl10	-0.32023			sp99.99	0.05	99.95
	1.94196			cm1 00	0.00	100.0
n3 Cl10	-0.31889			sp1.00	0.00	100.0
n1 Cl11	1.99412			cm0.19	94 7E	15.05
	-0.92090			sp0.18	84.75	15.25
	1.97382			~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	0.05	00.05
n2 Cl11	-0.32022			sp99.99	0.05	99.95
n3 Cl11	1.94195				0.00	100.0
	-0.31888			sp1.00	0.00	100.0
n1 O15	1.98601			am() 99	53.11	46.89
	-0.60657			sp0.88	55.11	40.89
m2 O15	1.96816			am1.00	0.00	100.0
n2 O15	-0.28535			sp1.00	0.00	100.0

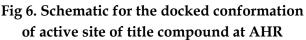
Table 3. NBO results showing the formation of Lewis and non-Lewis orbitals.

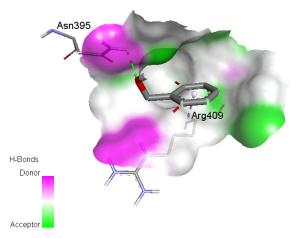
^a ED/e is expressed in a.u.

structure of aryl hydrocarbon receptor was downloaded from the protein data bank website (PDB ID: 2B02). All molecular docking calculations were performed on AutoDock-Vinasoftware [38]. The 3D crystal structure of aryl hydrocarbon receptor was obtained from Protein Data Bank. The protein was prepared for docking by removing the co-crystallized ligands, waters, and co-factors. The AutoDockTools (ADT) graphical user interface was used to calculate Kollman charges and polar hydrogen. The ligand was prepared for docking by minimizing its energy at B3LYP/6-31G (6D, 7F) level of theory. Partial charges were calculated by Geistenger method. The active site of the enzyme was defined to include residues of the active site within the grid size of 40Å×40Å×40Å. The most popular algorithm available in Auto Dock, the Lamarckian Genetic Algorithm (LGA), was employed for docking. The docking protocol was tested by extracting co-crystallized

inhibitor from the protein and then docking the same. The docking protocol predicted the same conformation as was present in the crystal structure with RMSD value well within the reliable range of 2Å [39]. Amongst the docked conformations, one which binds well at the active site was analyzed for detailed interactions in Discover Studio Visualizer 4.0 software. The ligand binds at the active site of the substrate (Figs. 6 and 7) by weak non-covalent interactions. Amino acid Asn395 forms H-bond with OH group and Arg409 forms hydrophobic interaction with phenyl ring. The docked ligand title compound forms a stable complex with aryl hydrocarbon receptor and gives a binding affinity (ΔG in kcal/mol) value of -4.4 (Table 4). These preliminary results suggest that the compound might exhibit inhibitory activity against aryl hydrocarbon receptor.







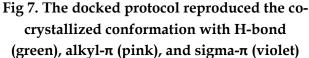


Table 4. The binding affinity values of different poses of the title compound predicted by AutoDockVina.

Mada	Affinity	Distance from best mode (Å)			
Mode	(kcal/mol)	RMSD l.b.	RMSD u.b.		
1	-4.4	0.000	0.000		
2	-4.3	13.604	14.299		
3	-4.3	13.610	14.145		
4	-4.3	0.643	2.985		
5	-4.2	21.000	22.118		
6	-4.2	1.948	4.008		
7	-4.1	2.155	2.445		
8	-4.1	12.995	13.858		
9	-4.1	2.144	2.938		

CONCLUSIONS

The vibrational spectroscopic studies of 2,6dichlorobenzyl alcohol were reported experimentally and theoretically. Potential energy distribution of normal modes of vibrations was done using GAR2PED program. Using HOMO and LUMO energy values, the quantum chemical descriptors are reported. MEP predicts the most reactive part in the molecule and it is evident that the negative region covers the CH₂ group, oxygen atom, and phenyl ring, and the positive region is over the hydrogen atoms. The hyperpolarizability of the title compound 4.523 times that of standard NLO material urea and is an attractive object for future studies in nonlinear optics. From the molecular docking study title compound forms a stable complex with aryl hydrocarbon receptor and gives a binding affinity value of -4.4kcal/mol, and this suggests that the compound might exhibit inhibitory activity against aryl hydrocarbon receptor.

ACKNOWLEDGEMENTS

Authors, BH would like to thank UGC, India for a minor research project and RKS would like thank University of Kerala for a research fellowship.

REFERENCES

- Schaefer T, Danchura W, Niemezura W, Parr WJE. Can. J. Chem., 1978, 56, 1721.
- Yu S, Andreichikov LA, Voronova ZD, Belykh AN, Perm Pharmaceutical Institute, USSR 666, 799, C1. C07D 267/14, 1979, p. 222.
- 3. Kenneth P, Albright JA, SCM Corp., 1973, p. 280.
- 4. Ogata H, Nawa H, Tokuda K, Ishihara M, Wako Pure Chemicals Industries, Ltd., 1987, p. 17.
- Porter DW, Bradley M, Brown Z, Charlton SJ, Cox B, Hunt P, Janus D, Lewis S, Oakley P, Connor DO, Reilly J, Smith N, Press NJ. *Bioorg. Med. Chem. Lett.*, 2014, 24, 3285.
- Serafini, MF. Testa MF, Villano D, Pecorari M, Wieren KV, Azzini E, Brambilla A, Maiani G. Free Radic Biol. Med., 2009, 15, 769.
- 7. Sundaraganesan N, Anand B, Meganathan C, Joshua BD, Saleem H. *Spectrochim. Acta*, 2008, 69, 198.
- 8. Sundaraganesan N, Anand B, Jian FF, Zhao P, *Spectrochim. Acta*, 2006, 65, 826.

- 9. Gaussian 09, Revision C.01, Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Mennucci B, Petersson GA, Nakatsuji H, Caricato M, Li X, Hratchian HP, Izmaylov AF, Bloino J, Zheng G, Sonnenberg JL, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery, Jr. JA, Peralta JE, OgliaroF, Bearpark M, Heyd JJ, Brothers E, Kudin KN, Staroverov VN, Keith T, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant JC, Iyengar SS, Tomasi J, Cossi M, Rega N, Millam JM, Klene M, Knox JE, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazvev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Martin RL, Morokuma K, Zakrzewski VG, Voth GA, Salvador P, Dannenberg JJ, Dapprich S, Daniels AD, Farkas O, Foresman JB, Ortiz JV, Cioslowski J, Fox DJ, Gaussian, Inc., Wallingford CT, 2010.
- 10. Becke AD. J. Chem. Phys. 1993, 98, 5648.
- 11. Lee C, Yang W, Parr RG, Phys. Rev. B. 1988, 37, 785.
- 12. Handy NC, Masley PE, Amos RD, Andrews JS, Murray CW, Laming G. *Chem. Phys. Lett.*, 1992, 197, 506.
- 13. Scott AP, Radom L. J. Phys. Chem., 1996, 100, 16502.
- Foresman JB in *Exploring Chemistry with Electronic* Structure Methods: A Guide to Using Gaussian, Ed. Frisch E, Pittsburg, PA, 1996.
- 15. Dennington R, Keith T, Millam J, *Gaussview, Version 5*, Semichem. Inc., Shawnee Missions, KS, 2009.
- 16. Martin JML, Van Alsenoy C, *GAR2PED*, *A Program to Obtain A Potential Energy Distribution from a Gaussian Archive Record*, University of Antwerp, Belgium, 2007.
- 17. N.P.G. Roeges NPG, A Guide to the Complete Interpretation of Infrared Spectra of Organic Structures, John Wiley and Sons Inc., New York, 1994.
- Colthup NB, Daly LH, Wiberly SE, Introduction to Infrared and Raman Spectroscopy, third ed., Academic Press, Boston, 1990.
- 19. Varsanyi G, Assignments of Vibrational Spectra of Seven hundred benzene derivatives, Wiley, New York, 1974.
- 20. Socrates G, *Infrared Characteristic Group Frequencies*, John Wiley and Sons, New York, 1981.
- 21. McMurry HL, Thornton V. Anal. Chem., 1952, 24, 310.
- 22. Varghese HT, Panicker CY, Philip D, Mannekutla JR, Inamdar SR. *Spectrochim. Acta*, 2007, 66, 959.
- 23. El-Shahway AS, Ahmed SM, Sayed NK. Spectrochim. Acta, 2007, 66, 143.
- 24. Mooney EF. Spectrochim. Acta, 1964, 20, 1021.
- 25. Mooney EF. Spectrochim. Acta, 1963, 19, 877.
- 26. Kleinman DA. Phys. Rev., 1962, 126, 1977.

- 27. Adant M, Dupuis M, Bredas JL. Int. J. Quantum. Chem., 1995, 56, 497.
- 28. Scrocco E, Tomasi J. Adv. Quantum. Chem., 1978, 103, 115.
- 29. Politzer P, Murray JS in: Theoretical Biochemistry and Molecular Biophysics: A Comprehensive Survey, Vol. 2, Protein, Beveridge DL, Lavery R, Eds., Adenine Press, Schenectady, NY, 1991, Chap. 13.
- 30. Scrocco E, Tomasi J. Top. Curr. Chem., 1973, 42, 95.
- 31. Parr RG, Szentpaly LV, Liu SJ. J. Am. Chem. Soc., 1999, 121, 1922.
- Chaltraj PK, Maiti B, Sarbar UJ. J. Phys. Chem., 2003, 107, 4973.
- 33. Parr RG, Donnelly RA, Levy M, Palke WE. J. Am. Chem. Soc., 1978, 68, 3801.
- 34. Parthasarathi R, Padmanabhan J, Subramanian V, Maiti B, Chattraj PK. J. Phys. Chem., 2003, 107, 10346.
- 35. Glendening ED, Reed AE, Carpenter JE, Weinhold F, *NBO 3.1 Program Manual*, Theoretical Chemistry Institute, University of Wisconsin, Madison, WI.
- Feng S, Cao Z, Wang X. BioChim. Biophys- A Rev, Cancer, 2013, 197, 1837.
- 37. Lawrence NJ, Hepworth LA, Denisson D, McGown AT, Hadfield JA. J. Fluorine Chem., 2003, 123, 101.
- 38. Trott O, Olson AJ. J. Comput. Chem., 2010, 31, 455.
- 39. Kramer B, Rarey M, Lengauer T. Proteins: Struct. Funct. Genet., 1999, 37, 228.